

# The Amidine Based Colorimetric Sensor for Fe<sup>3+</sup>, Fe<sup>2+</sup>, and Cu<sup>2+</sup> in Aqueous Medium

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**Abstract** An amidine based chemosensor **AM-1** was synthesized and characterized by various spectroscopic (FT-IR, <sup>1</sup>H-NMR and mass) data and elemental analyses. Sensor **AM-1** exhibited high selectivity and sensitivity towards Fe<sup>3+</sup>, Fe<sup>2+</sup> and Cu<sup>2+</sup> in the presence of other surveyed ions (such as Sr<sup>2+</sup>, Cr<sup>3+</sup>, Co<sup>2+</sup>, Ni<sup>2+</sup>, Zn<sup>2+</sup>, Ag<sup>+</sup>, Al<sup>3+</sup>, Ba<sup>2+</sup>, Ca<sup>2+</sup>, Cd<sup>2+</sup>, Cs<sup>+</sup>, Hg<sup>2+</sup>, K<sup>+</sup>, Li<sup>+</sup>, Mg<sup>2+</sup>, Mn<sup>2+</sup>, Na<sup>+</sup> and Pb<sup>2+</sup>) with a distinct naked-eye detectable color change and a shift in the absorption band. Moreover, the emission of **AM-1** was quenched selectively only in the presence of Fe<sup>3+</sup>.

**Keywords** Amidine · Chemosensor · Chromogenic receptor · Multi metal ion sensor · DFT

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## Introduction

Chemosensors are molecules of abiotic origin that reveal a significant change in the electronic, magnetic or optical signals when interacted with a specific guest/s (ions/molecules) [1, 2]. Among the various chemosensing systems, sensors based on a naked-eye response (colorimetric) have many advantages because of their ability to provide a simple, sensitive, selective, precise and economical method for the detection of a target analyte without the use of specific/sophisticated instrumentation [3–6]. In colorimetric chemosensors, the spectral and visual color changes i.e. the responses obtained at the molecular level in solution opened the door for the qualitative and quantitative determination of the target analyte.

Recently, the design and synthesis of selective and sensitive chemosensors for the selective detection of transition metal ions has gained enormous attention because of their participation in a variety of fundamental biological and physiological processes including in human metabolic processes [7–10]. As important physiologically associated metal ions, both the Fe<sup>2+</sup> and Fe<sup>3+</sup> ions play an indispensable role in many biochemical processes at the cellular level. Numerous enzymes use iron ions as a catalyst for electron transfer, oxygen metabolism, and RNA and DNA synthesis [11, 12]. However, both its deficiency (hypoferremia) and excess (hyperferremia) can induced a variety of diseases. Several serious diseases such as Alzheimer, Parkinson's and Huntington's disease etc. known to occur due to the cellular toxicity caused by iron ions [13, 14]. Similarly, Cu<sup>2+</sup> is a major metal pollutant because of its widespread use [15]. Cu<sup>2+</sup> ions play an important role as a catalytic cofactor in a variety of metallo-enzymes, including superoxide dismutase, cytochrome oxidase, and tyrosinase [16]. Cu<sup>2+</sup> also exhibits toxicity allied with neurological diseases such as Alzheimer's, Wilson's and prion disease [17–20]. Due to the biological importance of Cu<sup>2+</sup>, Fe<sup>2+</sup>, and

Fe<sup>3+</sup> ions, a method for the rapid, selective and sensitive recognition of these ions in food, pharmaceutical products and biological samples such as blood, urine, etc. is of great significance.

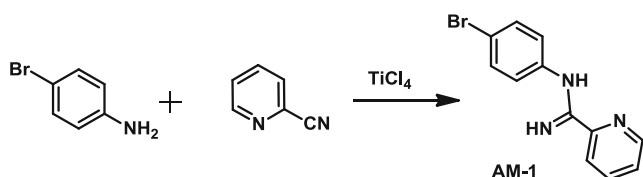
As a result, intense research has been focused on the development of sensitive and selective receptors for the qualitative and quantitative recognition of Cu<sup>2+</sup>, Fe<sup>2+</sup>, and Fe<sup>3+</sup>. Interestingly, as summarized in Table S1, the various reported sensors are quite specific, either for Cu<sup>2+</sup> or Fe<sup>2+</sup> or Fe<sup>3+</sup>, or for Cu<sup>2+</sup> and Fe<sup>3+</sup>, or for Fe<sup>2+</sup> and Fe<sup>3+</sup> [4, 5, 21–30], but to the best of our knowledge, a colorimetric chemosensor for the simultaneous detection of Cu<sup>2+</sup>, Fe<sup>2+</sup> and Fe<sup>3+</sup> ions remains unreported. Herein, as a part of our efforts in the field of analyte recognition [11, 29–33], the chemosensor **AM-1** has been developed for the selective and sensitive detection of Cu<sup>2+</sup>, Fe<sup>2+</sup> and Fe<sup>3+</sup> ions in aqueous medium. The cation recognition ability of **AM-1** towards different metal ions was studied by experimental (naked-eye, UV-visible, and fluorescence spectroscopy) and theoretical methods. Obtained results in naked eye study have successfully applied the receptor **AM-1** as an indicator for the ‘in situ’ qualitative detection of Fe<sup>3+</sup>, Fe<sup>2+</sup> and Cu<sup>2+</sup>. Interestingly, the fluorescence of receptor **AM-1** was quenched selectively in the presence of Fe<sup>3+</sup> among the other tested metal ions including Fe<sup>2+</sup> and Cu<sup>2+</sup>.

## Results and Discussion

The receptor **AM-1** was efficiently synthesized by the reaction of 2-cyanopyridine and 4-bromoaniline in the presence of TiCl<sub>4</sub> (Scheme 1) [34]. The molecular structure of **AM-1** was established by various spectroscopic (FT-IR, <sup>1</sup>HNMR and mass) data and elemental analyses (Fig. S1–3). The recognition ability of **AM-1** towards different metal ions was studied by naked eye, UV-visible and fluorescence spectroscopy. The obtained results were supported by theoretical study.

### UV–Vis Study

The UV–Vis absorption spectra of the receptor **AM-1** (5 × 10<sup>−5</sup> M, in methanol) was studied in the absence and presence of 5 equivalents of different metal ions such as Sr<sup>2+</sup>, Cr<sup>3+</sup>, Co<sup>2+</sup>, Ni<sup>2+</sup>, Zn<sup>2+</sup>, Ag<sup>+</sup>, Al<sup>3+</sup>, Ba<sup>2+</sup>, Ca<sup>2+</sup>, Cd<sup>2+</sup>, Cs<sup>+</sup>, Hg<sup>2+</sup>, K<sup>+</sup>,



**Scheme 1** Synthesis of receptor **AM-1**

Li<sup>+</sup>, Mg<sup>2+</sup>, Mn<sup>2+</sup>, Na<sup>+</sup>, Pb<sup>2+</sup>, Cu<sup>2+</sup>, Fe<sup>2+</sup> and Fe<sup>3+</sup> (1 × 10<sup>−2</sup> M, in water). Receptor **AM-1** exhibited two absorption bands at 225 and 272 nm. Upon addition of Fe<sup>3+</sup> ions to the solution of **AM-1**, a hypochromic shift was observed at 225 nm while a blue shift of 6 nm was observed at 272 nm with the appearance of a new broad charge transfer band between 350 and 450 nm. The charge transfer band of lower intensity was also observed in the presence Cu<sup>2+</sup> and Fe<sup>2+</sup> (Fig. 1). The spectral changes associated with **AM-1** can be attributed to the intermolecular charge transfer (ICT) process occurred due to the delocalization of electrons from the imine nitrogen (C=N) of the amidine to the metal ions during complexation. However, no significant changes in the absorption spectrum of **AM-1** were observed with other metal ions, there by revealing the selectivity towards Cu<sup>2+</sup>, Fe<sup>2+</sup> and Fe<sup>3+</sup>.

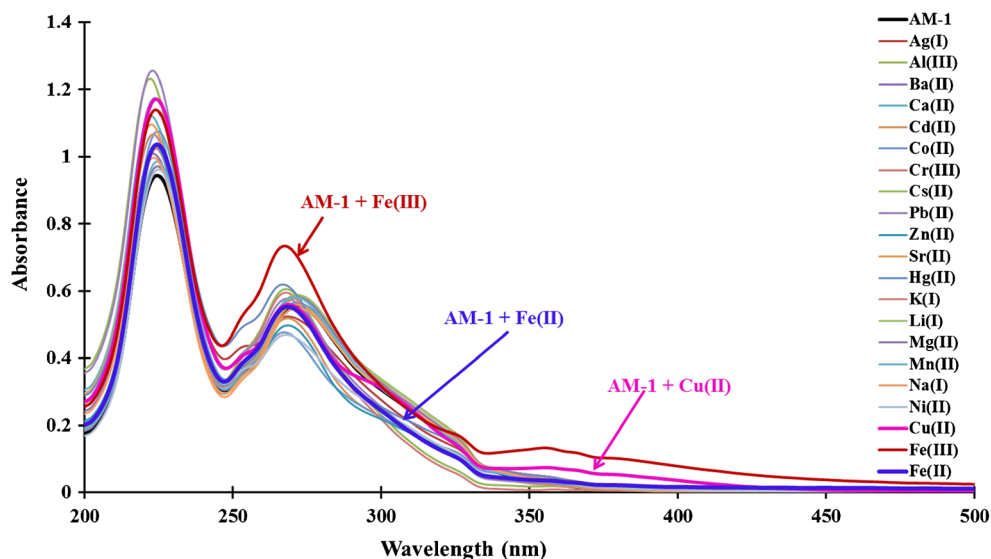
The absorption titrations of **AM-1** with the metal ions (Cu<sup>2+</sup>, Fe<sup>2+</sup>, and Fe<sup>3+</sup>) were next performed to examine the recognition ability. As shown in Fig. 2, on successive addition of incremental amounts of Fe<sup>3+</sup> to the **AM-1** solution, the receptor bands were gradually shifted with the appearance of a new charge transfer band. Using the absorption titration data, the binding constant of 5 × 10<sup>3</sup> M<sup>−1</sup> was calculated for Fe<sup>3+</sup> from the Benesi-Hildebrand Plot (Fig. S4). The detection and quantification limits of 0.6 and 1.9 μM, respectively for Fe<sup>3+</sup> were estimated (Fig. S5). Similarly, the binding constants of 5.2 × 10<sup>3</sup> M<sup>−1</sup> and 9.7 × 10<sup>2</sup> M<sup>−1</sup> were calculated for the metal ions Cu<sup>2+</sup> and Fe<sup>2+</sup> respectively from the Benesi-Hildebrand Plots (Fig. S6–7).

The binding stoichiometry for **AM-1**.Fe<sup>3+</sup>, **AM-1**.Fe<sup>2+</sup> and **AM-1**.Cu<sup>2+</sup> complexes were calculated by plotting the Job’s plot between the mole fractions of Fe<sup>3+</sup>, Fe<sup>2+</sup> and Cu<sup>2+</sup> with the absorption changes at 267 nm (Fig. 3). The maximum was obtained at a molar fraction of 0.5, which clearly delineated the formation of a ferric complex in 1:1 stoichiometry. Further, the stable **AM-1**.Fe<sup>2+</sup> and **AM-1**.Cu<sup>2+</sup> complexes were synthesized and characterized by LC-MS. The proposed 1:1 stoichiometry for both **AM-1**.Fe<sup>2+</sup> and **AM-1**.Cu<sup>2+</sup> complexes was supported by the obtained mass data (Fig. S8–9).

### Colorimetric (Naked Eye) Study

In colorimetric experiments, significant color changes of the **AM-1** solutions were observed upon addition of Fe<sup>3+</sup>, Fe<sup>2+</sup>, and Cu<sup>2+</sup> over the other tested metal ions (Fig. 4). A distinct color change of **AM-1** from colorless to red, orange and green was observed in the presence of Fe<sup>2+</sup>, Fe<sup>3+</sup> and Cu<sup>2+</sup> ions respectively, which indicates the sensitive and selective ‘naked-eye’ detecting ability for these cations. By utilizing the benefits of the ‘naked-eye’ results, we have applied the receptor **AM-1** as an indicator for the ‘in situ’ qualitative detection of Fe<sup>3+</sup>, Fe<sup>2+</sup> and Cu<sup>2+</sup> (Fig. 5). A solution of receptor **AM-1** (3 mL, 1 × 10<sup>−3</sup> M, in methanol) is colorless. After addition of 1 equivalent of Fe<sup>2+</sup>, it turns into dark blood red color. When

**Fig. 1** Absorbance spectral changes of **AM-1** (2 mL,  $5 \times 10^{-5}$  M, in methanol) upon addition of 5 equivalents of various metal ions (50  $\mu$ L,  $1 \times 10^{-2}$  M, in water)

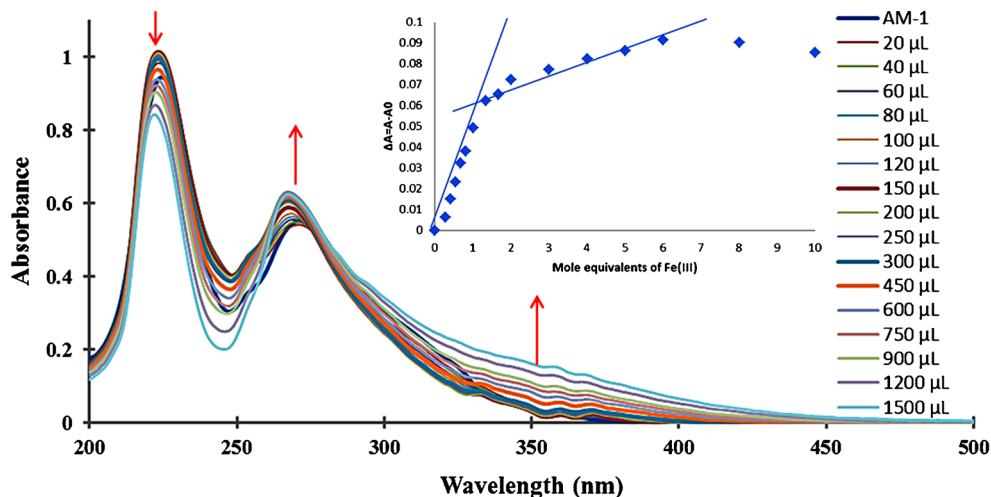


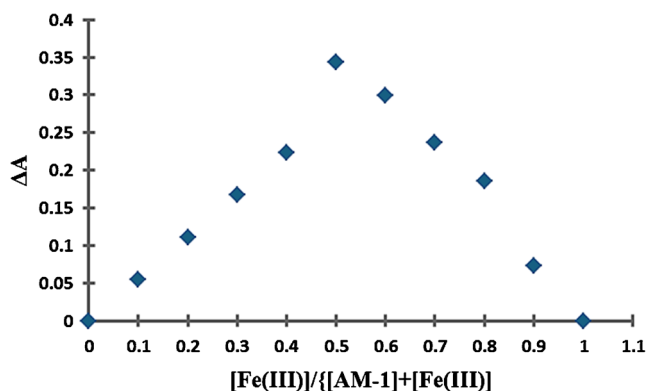
0.1 M  $K_2Cr_2O_7$  solution was added to this red color solution, immediate color change from dark red to faint yellow was observed which supports the conversion of  $Fe^{2+}$  into  $Fe^{3+}$ . On further addition of 0.1 M  $NaBH_4$ , the observed faint yellow color was converted reversibly into a dark red color. Further, the colorless solution of **AM-1** was reappeared with the excess addition of  $NaBH_4$ . These results inferred that we can monitored the presence of both the oxidation states of iron ( $Fe^{2+}$  and  $Fe^{3+}$ ) by using the receptor **AM-1**. In another experiment, we have added copper powder to the colorless solution of receptor **AM-1**. To this colorless mixture, addition of 0.1 M  $AgNO_3$  solution resulted in a remarkable color change from colorless to green, which indicates the conversion of metallic copper into the  $Cu^{2+}$  state. Moreover, it is interesting to mention here that all three ions ( $Fe^{2+}$ ,  $Fe^{3+}$  and  $Cu^{2+}$ ) shows reversibility in the color change on addition of aq. EDTA solution. The above naked-eye results opens the door for the applications of **AM-1** for the 'in situ' qualitative determination of three ions, namely  $Fe^{3+}$ ,  $Fe^{2+}$  and  $Cu^{2+}$ .

### Fluorescence Study

The cation binding behavior of **AM-1** was also investigated by emission measurements. Addition of  $Fe^{3+}$  ions (5 equivalents, 50  $\mu$ L,  $1 \times 10^{-2}$  M, in water) to the **AM-1** solution (2 mL,  $5 \times 10^{-5}$  M) in methanol results in a distinct fluorescence quenching (Fig. 6). However, in the presence of other metal ions such as  $Fe^{2+}$ ,  $Cu^{2+}$ ,  $Sr^{2+}$ ,  $Cr^{3+}$ ,  $Co^{2+}$ ,  $Ni^{2+}$ ,  $Zn^{2+}$ ,  $Ag^+$ ,  $Al^{3+}$ ,  $Ba^{2+}$ ,  $Ca^{2+}$ ,  $Cd^{2+}$ ,  $Cs^+$ ,  $Hg^{2+}$ ,  $K^+$ ,  $Li^+$ ,  $Mg^{2+}$ ,  $Mn^{2+}$ ,  $Na^+$  and  $Pb^{2+}$ , the fluorescence intensity of **AM-1** at 305 nm either did not induced any significant changes or was slightly enhanced under identical conditions. In another experiment, the  $Fe^{3+}$  sensing ability of **AM-1** under a competition environment was investigated in the presence of potentially interfering metal ions by fluorescence measurements (Fig. S10). For methanol solutions of **AM-1**, addition of 2 equiv. of  $Fe^{3+}$  in the presence of 2 equiv. of other tested metal ions caused a dramatic quenching in the fluorescence intensity of **AM-1** with either very slight or no interference effects. Therefore,

**Fig. 2** Absorption spectral changes of **AM-1** (3 mL,  $5 \times 10^{-5}$  M, in methanol) upon addition of 0–10 equivalents of  $Fe^{3+}$  ions (0–1,500  $\mu$ L,  $1 \times 10^{-3}$  M, in water). Inset: mole ratio plot from absorption titration of **AM-1** with  $Fe^{3+}$





**Fig. 3** Jobs Plot for AM-1 and  $\text{Fe}^{3+}$ , indicating the formation of a 1:1 (L:M) complex

it can be concluded that AM-1 is a reliable, highly selective and sensitive turn-off fluorescent sensor for  $\text{Fe}^{3+}$ .

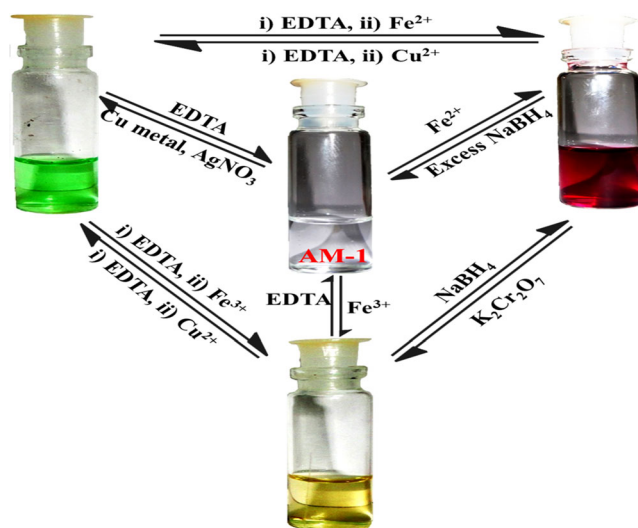
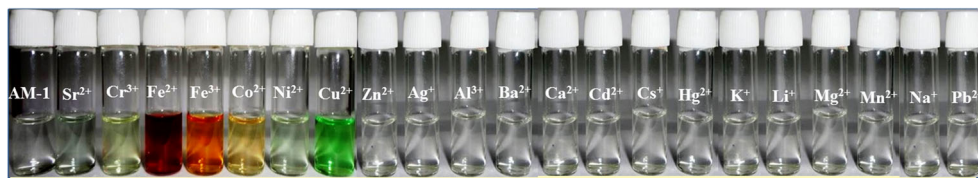
The fluorescence titration of AM-1 (2 mL,  $5 \times 10^{-5}$  M, in methanol) with  $\text{Fe}^{3+}$  ( $1 \times 10^{-3}$  M, in water) was performed via the successive addition of incremental concentrations of  $\text{Fe}^{3+}$  (0–1,000  $\mu\text{L}$ ). The fluorescence intensity of AM-1 was gradually quenched at 305 nm (Fig. 7). Based on the fluorescence titration data, the detection and quantification limits of 14 and 41  $\mu\text{M}$ , respectively for  $\text{Fe}^{3+}$  were estimated (Fig. S11). Also, the binding constant ( $K$ ) of AM-1 with  $\text{Fe}^{3+}$  was determined by a Benesi-Hildebrand plot (Fig. S12) and a Scatchard plot (Fig. S13) from the fluorescence titration data. The binding affinity of AM-1 was found to be  $\approx 1 \times 10^5 \text{ M}^{-1}$  for  $\text{Fe}^{3+}$ .

The quenching process can be mathematically expressed by the Stern–Volmer Eq. (1), which allows us to determine the types of quenching. If the Stern–Volmer plot is linear then the quenching is of a static type rather than dynamic quenching [35, 36].

$$F_0/F = 1 + k_q \tau_0 [Q] = 1 + K_{sv} [Q] \quad (1)$$

Where,  $F_0$  and  $F$  are the fluorescence intensities in the absence and presence of the quencher,  $k_q$  is the bimolecular quenching constant,  $\tau_0$  is the lifetime of the fluorescence in the absence of the quencher,  $[Q]$  is the concentration of the quencher, and  $K_{sv}$  is the Stern–Volmer quenching constant. In the presence of a quencher, the fluorescence intensity is reduced from  $F_0$  to  $F$ . Therefore, the ratio ( $F_0/F$ ) is directly proportional to the quencher concentration  $[Q]$ . According to Eq. (1), the Stern–Volmer plot of  $F_0/F$  versus  $[Q]$  shows a linear graph with a slope of  $K_{sv}$  (Fig. 8). The linear Stern–

**Fig. 4** Photo of the vials containing AM-1 (3 mL,  $1 \times 10^{-3}$  M, in methanol) in the presence of various metal ions (300  $\mu\text{L}$ ,  $1 \times 10^{-2}$  M, in water)

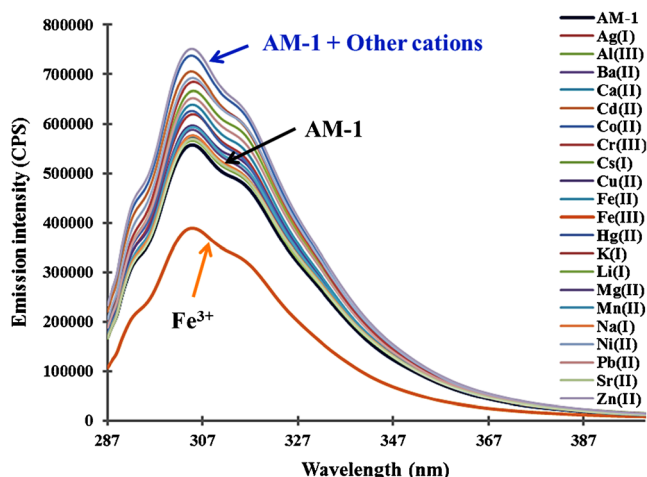


**Fig. 5** ‘In situ’ qualitative detection of  $\text{Fe}^{3+}$ ,  $\text{Fe}^{2+}$ , and  $\text{Cu}^{2+}$

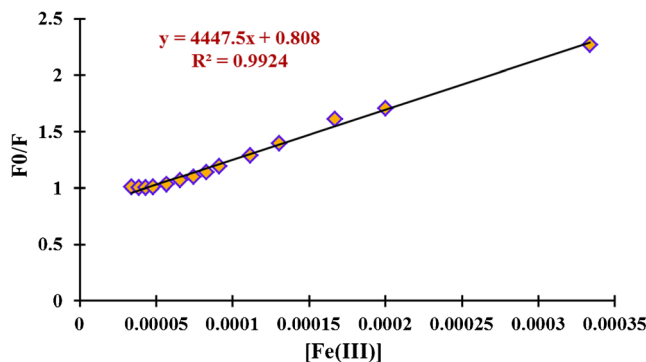
Volmer plot indicates that the quenching is static for the AM-1. $\text{Fe}^{3+}$  system [35]. The results also confirmed the formation of only one type of complex between the receptor AM-1 and the  $\text{Fe}^{3+}$  ions.

#### Computational Study

In the absence of a suitable single crystal of AM-1 and its complexes, and to get more insight into the above experimental observations, DFT calculations were performed to understand the electronic environment and the changes in the structure of AM-1 upon complexation with  $\text{Fe}^{3+}$ ,  $\text{Fe}^{2+}$  and  $\text{Cu}^{2+}$ . During the computational study, two stable conformations of AM-1 (L and L') were optimized (Fig. 9). The form L is relatively more stable than L' by 5.01 kcal/mol. There is an intramolecular H-bond (2.036 Å) between the amine proton (H6) with the pyridine-N atom. Also, the ligand exhibited two possible coordination modes (Fig. 10, mode I: Py-N and =NH and mode II: Py-N and -NH) forming a five-membered chelated ring with the metal ions. According to the calculated relative energy, the complexes preferred to coordinate through the Py-N and =NH. The  $\text{Cu}^{2+}$ ,  $\text{Fe}^{2+}$  and  $\text{Fe}^{3+}$  complexes with the coordination mode I is relatively more stable than mode II by 25.15, 7.36 and 54.32 kcal/mol, respectively. In coordination mode I, the interaction energy was calculated by applying the equation  $[E_{\text{int}} = E_{(\text{ML})} - E_{(\text{L})} - E_{(\text{M})}]$ . The  $E_{\text{int}}$  for the AM-1. $\text{Fe}^{3+}$ , AM-1. $\text{Fe}^{2+}$  and AM-1. $\text{Cu}^{2+}$  complexes are  $-792.58$ ,  $-286.83$  and  $-330.29$  kcal/mol, respectively. This result



**Fig. 6** Fluorescence spectral ( $\lambda_{ex}=277$  nm) changes of AM-1 ( $5 \times 10^{-5}$  M, in methanol) upon addition of 5 equivalent of various metal ions ( $1 \times 10^{-2}$  M, in water)



**Fig. 8** Stern-Volmer Quenching Plot for Fe<sup>3+</sup> with receptor AM-1

indicates that Fe<sup>3+</sup> is forming a stronger complex with AM-1 followed by Cu<sup>2+</sup> and then Fe<sup>2+</sup>.

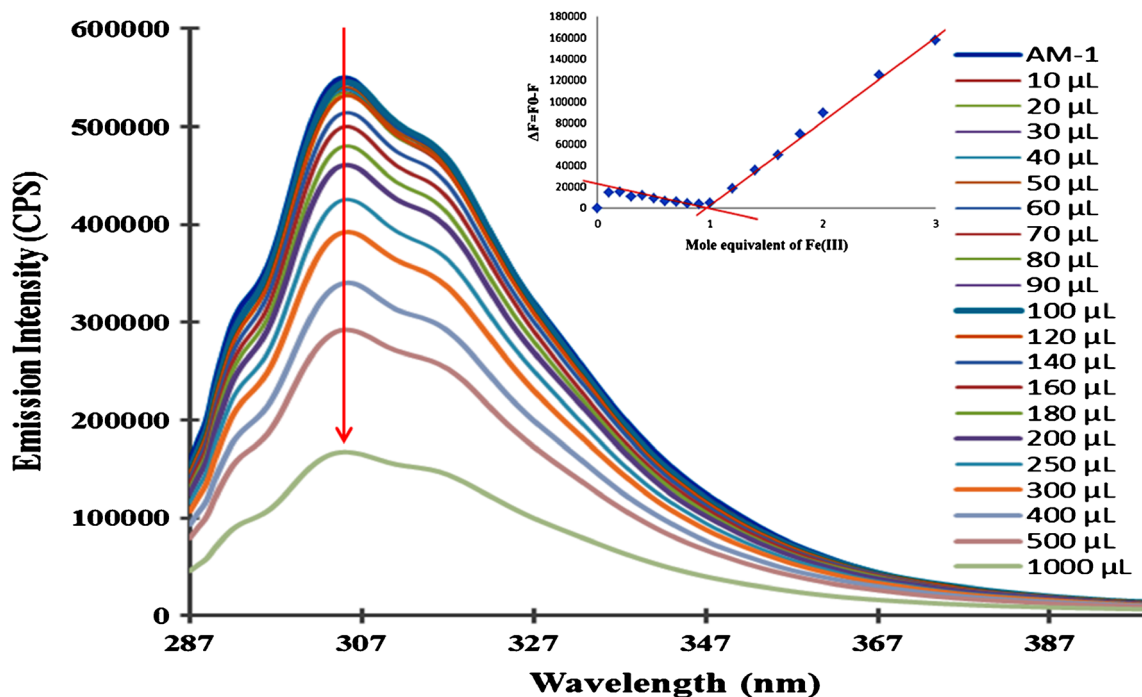
The plots of the frontier molecular orbital's (FMO's) of AM-1 and its complexes with Fe<sup>3+</sup>, Cu<sup>2+</sup> and Fe<sup>2+</sup> were analyzed. As shown in Fig. S14 and S15, the electron density of the highest-occupied molecular orbital (HOMO) and lowest-unoccupied molecular orbital (LUMO) of the receptor located in two different rings suggests a strong intramolecular charge transfer. On complexation, the lowering of the band gap indicates

the shift in the absorption band, and simultaneously the HOMO and LUMO indicates the formation of a charge-transfer complex between the receptor and metal ions.

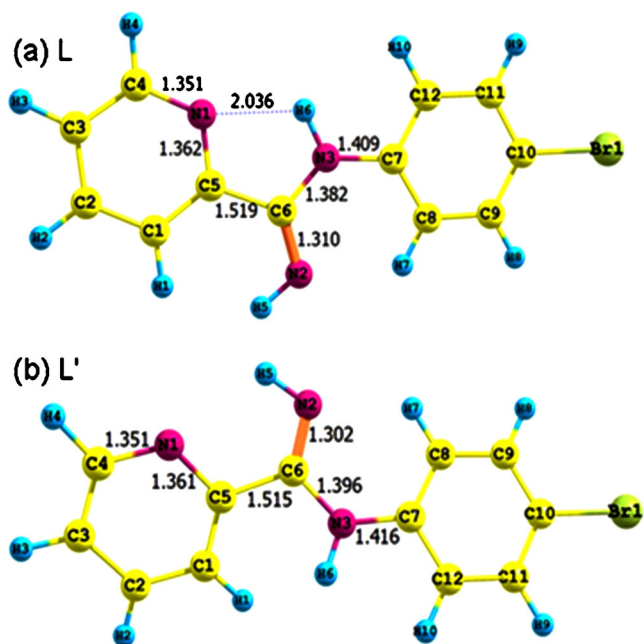
### Experimental

#### Materials and Methods

All the starting reagents and metal perchlorates were purchased either from S. D. Fine chemicals or Sigma Aldrich depending on their availability. All the reagents were used as received. All the solvents were of spectroscopic grade and were used without further treatment. The purity of the compounds and the progress of reactions were determined and



**Fig. 7** Fluorescence spectral changes of sensor AM-1 (2 mL,  $5 \times 10^{-5}$  M, in methanol) upon addition of 0–10 equivalents of Fe<sup>3+</sup> (0–1,000 μL,  $1 \times 10^{-3}$  M, in water) at  $\lambda_{ex}=277$  nm. *Inset:* mole ratio plot from fluorescence titration of AM-1 with Fe<sup>3+</sup>



**Fig. 9** Optimized structure of the two probable conformations (L and L') of AM-1 at B3LYP/SDD method

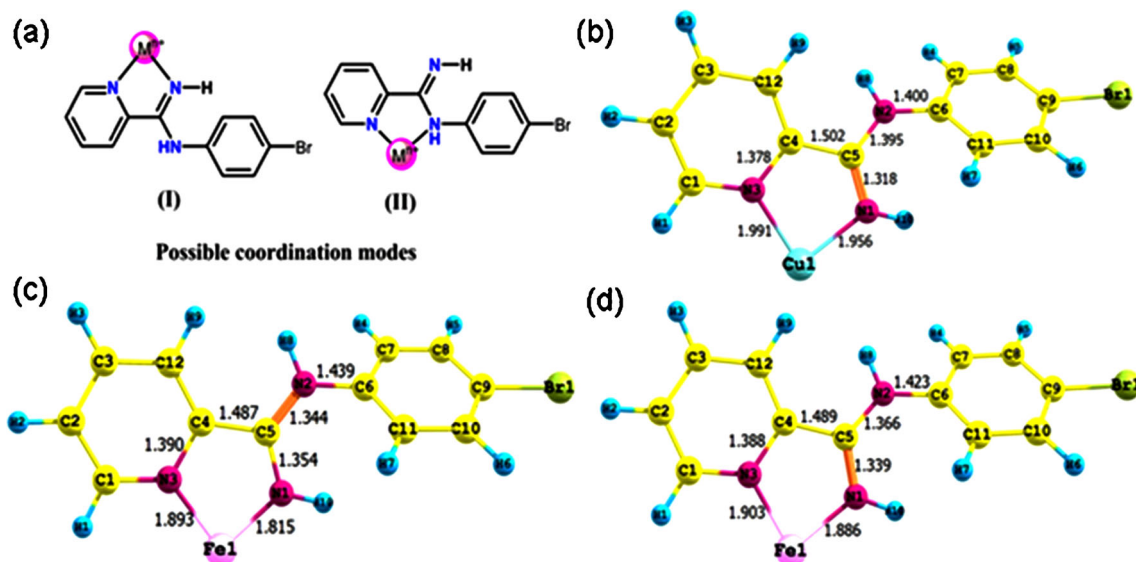
monitored by means of analytical thin layer chromatography (TLC). Pre-coated silica gel 60 F<sub>254</sub> (Merck) on alumina plates (7×3 cm) were used and visualized by using either an iodine chamber or a short UV-Visible lamp. Melting points were recorded on the Celsius scale by open capillary method and are uncorrected. IR spectra were recorded on a Perkin-Elmer Spectrum One FT-IR spectrometer as potassium bromide pellets and nujol mulls, unless otherwise mentioned. NMR spectra were recorded in CDCl<sub>3</sub> on a Varian (Mercury Vx) SWBB Multinuclear probe spectrometer, operating at 300 and 75 MHz for <sup>1</sup>H NMR and <sup>13</sup>C NMR, respectively and shifts

are given in ppm downfield from tetramethylsilane (TMS) as an internal standard. UV-Vis spectra were recorded on a U-3900 spectrophotometer (Perkin Elmer Co., USA) with a quartz cuvette (path length=1 cm). Fluorescence spectra were recorded on a Fluoromax-4 spectrofluorometer (HORIBA Jobin Yvon Co., France).

#### Synthesis of AM-1

In 250 ml dry round bottom flask, a mixture of 4-bromoaniline (1.28 g, 10 mmol) and 2-cyanopyridine (1.04 g, 10 mmol) was heated in an oil bath at a temperature range of 110–120 °C with constant stirring. After 30 min., TiCl<sub>4</sub> (1.33 ml, 12 mmol) was added to the flask. Then, the temperature was increased to 150–160 °C, and heating was continued for 3–4 h. The progress of reaction was monitored by TLC. After completion of the reaction, the obtained solid was cooled to room temperature and dissolved in a hot water followed by the solution was made alkaline with 10 % NaOH solution. This alkaline solution was extracted with dichloromethane [3×50 mL]. The organic layer was decolorized with activated charcoal and dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. After evaporating the solvent under reduced pressure, the crude amidine was obtained. This crude amidine was recrystallized from an acetone:hexane [10:90] system, to afford pure amidine. Yield: 2.24 g (81 %).

**IR [KBr, cm<sup>-1</sup>]:** 3,376, 2,854, 1,640, 1,577, 1,530, 1,459, 1,377, 1,321, 1,300, 1,260, 1,151, 1,119, 1,096, 1,006, 818, 769, 722, 541. **LCMS [ESI, e/z (%)] :** 278 (100), 276 (100), 260 (20), 259 (20). **<sup>1</sup>H NMR [CDCl<sub>3</sub>, 300 MHz]:** 5.85 (br s, 2H, NH, C=NH), 6.88–6.91 (d, J=8.7 Hz, 2H, ArH), 7.38–7.50 (m, 3H, ArH), 7.79–7.84 (m, 1H, ArH), 8.36–8.39 (d, J=8.1 Hz, 1H, ArH), 8.56–8.58 (m, 1H, ArH). Anal. calcd for



**Fig. 10** a The possible coordination modes of AM-1, and the favorable optimized structure of the complexes **b** AM-1.Cu<sup>2+</sup>, **c** AM-1.Fe<sup>2+</sup> and **d** AM-1.Fe<sup>3+</sup> at B3LYP/SDD method [36]

C<sub>12</sub>H<sub>10</sub>BrN<sub>3</sub>: C, 52.20; H, 3.65; N, 15.22. Found: C, 52.54; H, 3.60; N, 15.28.

### Spectroscopic Study

The aqueous stock solutions of cations of concentration  $1 \times 10^{-2}$  mol L<sup>-1</sup> were prepared from their corresponding salt i.e. AgClO<sub>4</sub>, Al(ClO<sub>4</sub>)<sub>3</sub>·9H<sub>2</sub>O, Ba(ClO<sub>4</sub>)<sub>2</sub>, Ca(NO<sub>3</sub>)<sub>2</sub>·4H<sub>2</sub>O, Cd(ClO<sub>4</sub>)<sub>2</sub>·H<sub>2</sub>O, Co(ClO<sub>4</sub>)<sub>2</sub>·6H<sub>2</sub>O, Cr(ClO<sub>4</sub>)<sub>3</sub>·6H<sub>2</sub>O, Fe(ClO<sub>4</sub>)<sub>2</sub>·xH<sub>2</sub>O, HgCl<sub>2</sub>, LiBr, Mg(ClO<sub>4</sub>)<sub>2</sub>, Mn(ClO<sub>4</sub>)<sub>2</sub>·H<sub>2</sub>O, Ni(ClO<sub>4</sub>)<sub>2</sub>·6H<sub>2</sub>O, Fe(ClO<sub>4</sub>)<sub>3</sub>·H<sub>2</sub>O, Pb(ClO<sub>4</sub>)<sub>2</sub>·3H<sub>2</sub>O, Zn(ClO<sub>4</sub>)<sub>2</sub>·6H<sub>2</sub>O, Cu(ClO<sub>4</sub>)<sub>2</sub>·6H<sub>2</sub>O, respectively. The stock solution of **AM-1** ( $1 \times 10^{-3}$  mol L<sup>-1</sup>) was prepared by dissolving an accurately weighed **AM-1** in CH<sub>3</sub>OH and then diluted to  $5 \times 10^{-5}$  mol L<sup>-1</sup> with CH<sub>3</sub>OH. These solutions were used for all spectroscopic studies after appropriate dilution. For spectroscopic titrations, the required amount of the diluted receptor **AM-1** was taken directly into a cuvette and the spectra were recorded after successive addition of the cations by using a micropipette.

### Computational Analysis

The structural optimization of the receptor and its complexes with the metal ions (Cu<sup>2+</sup>, Fe<sup>2+</sup> and Fe<sup>3+</sup>) was performed by applying the density functional theory (DFT) method in the gas phase by using the computer program Gaussian 09W [36]. All DFT calculations were performed with a hybrid functional B3LYP (Becke's three parameter hybrid functional using the LYP correlation functional) using the basis sets SDD. Then, the optimized geometries have been confirmed by frequency analyses at the same level of theory to ascertain the optimized structure were stable.

### Conclusion

In summary, we have developed an easy-to-synthesize amidine based receptor **AM-1** for the selective detection of Fe<sup>2+</sup>, Fe<sup>3+</sup> and Cu<sup>2+</sup> ions by colorimetric and UV–Vis spectral changes. The 1:1 binding stoichiometry for the **AM-1**·Fe<sup>3+</sup>, **AM-1**·Fe<sup>2+</sup>, and **AM-1**·Cu<sup>2+</sup> complexes was proposed by Job's plot and LC-MS analysis. Interestingly, the receptor exhibited a highly selective and sensitive fluorescence turn-off response only in the presence of Fe<sup>3+</sup>. Based on the spectral responses, the detection and quantification limits for receptor **AM-1** for Fe<sup>3+</sup> were estimated at 0.6 and 1.9 μM, respectively from absorption measurements, whereas values of 14 and 41 μM, respectively were obtained from emission measurements. The sensor **AM-1** opens the door for the applications of **AM-1** for the 'in situ' qualitative determination of three ions, namely Fe<sup>3+</sup>, Fe<sup>2+</sup> and Cu<sup>2+</sup>. The receptor **AM-1** with

its' low cost, ease of preparation and impressive selectivity inferred that this approach could potentially lead to many more sensors being designed using the amidine moiety as a core skeleton.

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